

Natural Products

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Aplysiasecosterol A: A 9,11-Secosteroid with an Unprecedented Tricyclic γ-Diketone Structure from the Sea Hare *Aplysia kurodai***

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Abstract: A new 9,11-secosteroid having an unprecedented tricyclic γ -diketone structure, aplysiasecosterol A (1), was isolated from the sea hare Aplysia kurodai. The structure was determined by one- and two-dimensional NMR spectroscopic analysis, molecular modeling studies, a comparison of experimental and calculated ECD spectra, and a modified Mosher's method. Aplysiasecosterol A (1) exhibited cytotoxicity against human myelocytic leukemia HL-60 cells. A biosynthetic pathway for 1 from a known cholesterol was proposed and includes twice α -ketol rearrangements and an intramolecular acetalization.

Sea hares (family Aplysiidae) belong to the opisthobranch group of mollusks (clade Aplysiomorpha).[1] They are shellless and benthic marine invertebrates, and have been postulated to contain chemical defense substances. Among them, the genera Aplysia and Dolabella are known to be rich sources of bioactive molecules.^[2] For example, dolastatin 10 is an antineoplastic peptide which was obtained from the Indian Ocean sea hare Dolabella sp.[3] In 2011, an mAb-targeted dolastatin 10 conjugate was approved for the treatment of Hodgkin's lymphoma. Aplyronine A, a highly potent antitumor and actin-depolymerizing macrolide, was isolated from the Japanese sea hare Aplysia kurodai.[4] Recently, aplyronine A was shown to induce protein-protein interactions between actin and tubulin and to prevent spindle formation and mitosis.^[5] In addition, various secondary metabolites have been isolated from A. kurodai, including the shikimate derivatives pericosines, [6] the sterol derivatives aplykurodins,^[7] and the alkaloids gliocladins^[8] and aplaminal.^[9] These molecules are expected to give new insights for the discovery and development of a new class of pharmacological tools and therapeutic agents.

In our continuing search for new bioactive compounds from *A. kurodai*, aplysiasecosterol A (1), a 9,11-secosteroid having an unprecedented tricyclic γ -diketone structure, was

Aplysiasecosterol A (1)

isolated. We report herein the isolation, structure determination, and bioactivity of 1.

The sea hare *A. kurodai* (54.8 kg, wet wt.) was extracted with aqueous ethanol. The concentrated extract was partitioned between ethyl acetate and water. The ethyl acetate layer was further partitioned with *n*-hexane, dichloromethane, and 60% aqueous methanol. Apart from the highly cytotoxic fractions which contain aplyronines, we investigated the other constituents in the dichloromethane layer. Repeated SiO₂, Al₂O₃, and ODS column chromatography, and final reverse-phase HPLC purification afforded aplysiasecosterol A (1) as a colorless oil {6.7 mg, 1.2×10^{-7} %, $[\alpha]_D^{24} = +19.3$ (*c* 0.228, MeOH)}. Aplysiasecosterol A (1) did not show significant cytotoxicity against the human cervical carcinoma cell line HeLa S3 at 200 μM, but it exhibited moderate cytotoxicity against the human myelomonocytic leukemia cell line HL-60 ($IC_{50} = 16 \mu M$).

The molecular formula of **1** was established to be $C_{27}H_{44}O_7$ by HR-ESIMS ([M+Na]⁺, m/z 503.2979, Δ –0.6 mmu). The planar structure of **1** was determined by one- and two-dimensional NMR analysis (Figure 1). The ¹H, ¹³C NMR, DEPT135, and HSQC spectra in CDCl₃ showed that **1** had the following signals: four singlets representing methyl groups (δ_H =0.89, 1.11, 1.21, 1.15 ppm), one doublet

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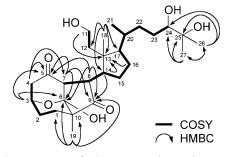


Figure 1. Planar structure of aplysiasecosterol A (1) determined by two-dimensional NMR analysis (bold line, COSY; arrows, selected HMBC correlations).

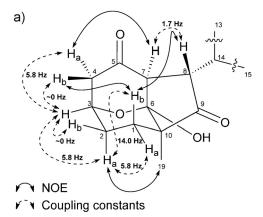


representing a methyl group ($\delta_{\rm H}\!=\!0.92$ ppm), two carbonyl carbon atoms ($\delta_{\rm C}\!=\!208.28,\ 216.50$ ppm), and one acetal carbon atom ($\delta_{\rm C}\!=\!101.31$ ppm). The IR (CHCl₃) spectrum of 1 showed absorption bands for hydroxy groups (3568 cm⁻¹) and two carbonyl groups (1736, 1708 cm⁻¹).

A detailed analysis of the COSY spectrum of 1 allowed us to construct three partial structures: C1-C4, C7-C24, and C11–C12 (Figure 1, Table S1). HMBC correlations between Me-18/C12, C13, C14, and C17 revealed that the C7-C24 unit was linked to the C11–C12 unit via the sp³ quaternary carbon C13. Similarly, HMBC correlations between Me-19/C1, C6, C9, and C10 indicated that C1, C6, C9, and C19 were each connected to the sp³ quaternary carbon C10. The C8–C9 connectivity was established based on HMBC correlations between H7, H8, and H14/C9. Further HMBC correlations between H7 and H8/C5 and C6 indicated that the carbonyl carbon C5 and the acetal carbon C6 were connected to the methine carbon C7. Thus, the results showed that 1 possesses a cyclopentanone moiety (C9, 1736 cm⁻¹) with an acetal carbon atom. Moreover, the presence of both a tetrahydropyran ring and a 4-oxacyclohexanone moiety (C5, 1708 cm⁻¹) in 1 was established by HMBC correlations between H3/C5 and C6 and H4/C5 and C7. Furthermore, the connectivity between two single methyl groups (C26, 27) and an 1,2-diol moiety was clarified based on HMBC correlations between Me-26/C24, C25, and C27, and Me-27/C24, C25, and C26. Based on the molecular formula and the degree of unsaturation, 1 was shown to contain four hydroxy groups. Therefore, the planar structure of 1 was determined to be as shown in

Next, the relative stereochemistry of the tricyclic γ -diketone structure of **1** was established. The large magnitude of $J_{1b,2a} = 14.0 \, \text{Hz}$ and the relatively small magnitudes of $J_{1a,2a} = 5.8 \, \text{Hz}$, $J_{2a,3} = 5.8 \, \text{Hz}$, and $J_{2b,3} = 0 \, \text{Hz}$ suggested that H1b and H2a were oriented in an *anti*-arrangement in the tetrahydropyran ring (Figure 2a). Key NOEs were observed for H1b/H4b and H1b/H8, and indicated that these three protons faced each other on the concave face of the rigid tricyclic structure. Similarly, NOE correlations for H4a/H7 and H2a/Me-19 suggested that all of the protons H2a, H4a, H7, and Me-19 were located in the convex face. Thus, H1b, H2a, and Me-19 were thought to be oriented in an axial position with respect to the tetrahydropyran ring in a chair conformation.

A molecular modeling study using a Merck molecular force field 94x (MMFF94x) showed that the tricyclic γ-diketone model compound **2**, in which the substituent on C8 was replaced with an isopropyl group, had only one conformer within 7 kcal mol⁻¹ of the lowest-energy conformation. Geometry optimization with the density functional theory (DFT) method for **2** was conducted using the B3LYP/6-31G + level of theory (Figure 2b). The calculated distances of H1b/H4b, H1b/H8, H2a/Me-19, and H4a/H7 for **2** were 3.1, 2.9, 2.5, and 3.1 Å, respectively. Thus, this model satisfied all of the key NOEs observed for **1**. The dihedral angles for H2b/H3 and H7/H8 in **2** were -67.8° and -105.8°, respectively. Based on the Karplus equation, [10] the coupling constants of H2b/H3 and H7/H8 in **2** were estimated to be 0–2 Hz, which coincided with those observed in **1** (ca. 0 and 1.7 Hz, respectively). For



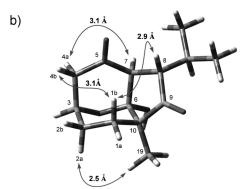


Figure 2. a) Relative stereochemistry of the tricyclic γ-diketone structure of 1 determined by one- and two-dimensional NMR analysis (solid arrows, selected NOE correlations; dashed arrows, coupling constants). b) Optimized structure of the tricyclic model compound 2 at the B3LYP/6-31G+ level of theory in the gas phase. The substituent on C8 in 2 was replaced with an isopropyl group. Solid arrows are the calculated distances between two selected protons.

the above reason, the relative stereochemistry of the tricyclic γ -diketone portion of **1** was determined to be as shown for the model compound **2** in Figure 2.

The relative stereochemistry around the cyclopentane ring in **1** was also determined by NOE experiments and coupling constant analysis (Figure 3). NOEs were observed for H11a/H14, H14/H17, H17/Me-21, H12a/Me-21, H16a/H22a, and Me-18/H20. These data strongly indicated that H12, H14, H17, and Me-21 are oriented in the one face, and Me-18 and H20 are oriented in the other face of the

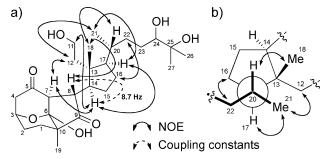


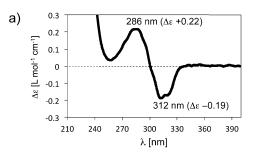
Figure 3. a) Relative stereochemistry around the cyclopentane ring of 1 determined by one- and two-dimensional NMR analysis (solid arrows, selected NOE correlations; dashed arrows, coupling constants). b) A Newman projection with a view along the C20–C17 bond.



cyclopentane ring. As a result, the relative stereochemistry of the cyclopentane ring part in 1 was identical to those of typical 9,11-secosteroids.[11,12]

The relative stereochemistry between the tricyclic structure and cyclopentane ring part in 1 was also determined based on a detailed NMR analysis. While the C8(sp³)-C14(sp³) bond is able to freely rotate, the large magnitude of $J_{8.14} = 8.7$ Hz indicated that H8 is positioned antiperiplanar to H14. NOEs were observed for H7/H14, H7/H12, and H8/Me-18. These results suggested that H7, H12, and H14 are oriented in the α -face, and H8 and Me-18 are oriented on β face of the cyclopentane ring. Thus, the relative stereochemistry of 1 except for the oxymethine carbon C24 was established to be as shown in Figure 3.

To determine the absolute stereochemistry of 1, the experimental electronic circular dichroism (ECD) spectrum of 1 was compared with the calculated ECD data for the model compound 2 (Figure 4).[13] The ECD spectrum of



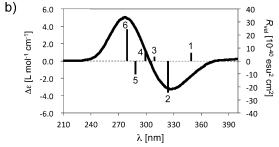


Figure 4. a) Experimental ECD spectrum of 1 measured in MeOH at 0.73 mm. b) Calculated ECD data for the model compound 2 at the PBEPBE/6-311G + + (d,p) level of theory in implicit solvent model (CPCM, MeOH) (solid line, left scale). Six first excited states for 2 were shown in solid bars (right scale).

1 showed a negative first Cotton effect at $\lambda = 312 \text{ nm}$ ($\Delta \varepsilon$ -0.19) and a positive second Cotton effect at $\lambda = 286$ nm ($\Delta \varepsilon$ +0.22). The calculated ECD spectrum for **2** at the PBEPBE/ 6-311G++(d,p) level of theory in implicit solvation model (CPCM, [14] MeOH) also had a negative ECD peak at $\lambda =$ 327 nm ($\Delta \varepsilon$ –3.3) and a positive ECD peak at λ = 277 nm ($\Delta \varepsilon$ + 5.0), which reproduced the signs of the experimental Cotton effects. As inferred from the molecular orbital analysis, the negative ECD band (band 2, $R_{\rm vel}$ -24.3) was mainly ascribed to the $n \rightarrow \pi^*$ transition of the C9 ketone (HOMO \rightarrow LUMO+1; Figure 5). Meanwhile, slightly positive ECD bands at $\lambda =$ 309 nm (band 3, $R_{\text{vel}} + 3.3$) and $\lambda = 299$ nm (band 4, $R_{\text{vel}} + 7.4$) were mainly ascribed to the $n \rightarrow \pi^*$ transition of the C5 ketone (HOMO-1→LUMO and HOMO-1→LUMO+1, respec-

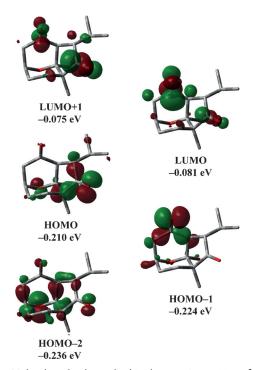


Figure 5. Molecular orbitals involved in the $n\rightarrow\pi^*$ transition of the C5 and C9 ketones in the model compound 2.

tively). In addition, the positive ECD band (band 6, R_{vel} +23.8) corresponded to the transition from HOMO-2 to LUMO + 1. These results suggested that the absolute configuration of the tricyclic γ -diketone part of $\mathbf{1}$ was identical to that of the model compound **2**.

Finally, 1 was converted into an 11,24-(MTPA) ester (see Figure S2 in the Supporting Information). Through the use of a modified Mosher's method, [15] the stereochemistry of C24 was determined to be R. For the above results, the absolute stereochemistry of 1 was completely established.

A biosynthetic pathway for the tricyclic γ-diketone structure of 1 was proposed, as shown in Scheme 1. Because of the structural similarity of the cyclopentane ring and the side-chain part of 1 with those of known 9,11-secosteroids, cholest-7-en-3S,5R,6R-triol^[16] was assumed to be a biosynthetic precursor of 1. Oxidative cleavage of the C9-C11 bond and oxidation of the C6 hydroxy group would give 1,4-diketone 3. The α -ketol rearrangement^[17] in 3 would lead to the formation of the C6–C10 bond to give the α , β -unsaturated ketone 4. The vinylogous α -ketol rearrangement in 4 might form the C5-C7 bond, and subsequent enolization at the C9 ketone would afford the enol 5. Finally, protonation at C8 and intramolecular acetalization of 5 would afford the tricyclic γdiketone structure of 1. The stereochemistry of the methine carbon at C8 in general steroids is opposite to that in 1. However, protonation at C8 in 5 might occur to give priority to the thermodynamically stable exo-substituent 8S isomer.

In summary, the structure and bioactivity of aplysiasecosterol A (1), a 9,11-secosteroid with a novel tricyclic γ diketone structure, was established. Structurally and functionally diverse secosteroids have been discovered from both terrestrial and marine origin, which include 5,6-, 8,9-, 8,14-,



Cholest-7-en-3S,5R,6R-triol

Scheme 1. Proposed biosynthetic pathway for the tricyclic γ -diketone structure of 1.

9,10-, 9,11-, and 13,17-secosteroids. [12] To the best of our knowledge, however, there are no examples having tricyclic ring systems similar to that of **1**. Further biological and biosynthetic studies on **1** are in progress.

Keywords: biosynthesis · circular dichroism · natural products · rearrangements · structure elucidation

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[1] T. E. Thompson, *Biology of opisthobranch molluscs, Vol. 1*, The Ray Society, London, **1976**.

- [2] a) M. Kita, H. Kigoshi, in *Handbook of Anticancer Drugs from Marine Origin* (Ed. S. K. Kim), Springer International Publishing, Switzerland, 2015, pp. 701–740.
- [3] G. R. Pettit, in *International Oncology Updates: Marine Anti-cancer Compounds in the Era of Targeted Therapies* (Eds. B. Chabner, H. Cortes-Funes), Permanyer, Barcelona, 2009, pp. 19–50.
- [4] a) K. Yamada, M. Ojika, T. Ishigaki, Y. Yoshida, H. Ekimoto, M. Arakawa, J. Am. Chem. Soc. 1993, 115, 11020-11021; b) M. Ojika, H. Kigoshi, T. Ishigaki, I. Tsukada, T. Tsuboi, T. Ogawa, K. Yamada, J. Am. Chem. Soc. 1994, 116, 7441-7442; c) K. Yamada, M. Ojika, H. Kigoshi, K. Suenaga, Nat. Prod. Rep. 2009, 26, 27-43.
- [5] a) M. Kita, Y. Hirayama, K. Yoneda, K. Yamagishi, T. Chinen, T. Usui, E. Sumiya, M. Uesugi, H. Kigoshi, J. Am. Chem. Soc. 2013, 135, 18089 18095; b) M. Kita, H. Kigoshi, Nat. Prod. Rep. 2015, 32, 534 542.
- [6] a) A. Numata, M. Iritani, T. Yamada, K. Minoura, E. Matsumura, T. Yamori, T. Tsuruo, *Tetrahedron Lett.* 1997, 38, 8215–8218; b) T. Yamada, M. Iritani, H. Ohishi, K. Tanaka, K. Minoura, M. Doi, A. Numata, *Org. Biomol. Chem.* 2007, 5, 3979–3986.
- [7] T. Miyamoto, R. Higuchi, T. Komori, T. Fujioka, K. Mihashi, Tetrahedron Lett. 1986, 27, 1153–1156.
- [8] Y. Usami, J. Yamaguchi, A. Numata, Heterocycles 2004, 63, 1123-1129.
- [9] T. Kuroda, H. Kigoshi, Org. Lett. 2008, 10, 489-491.
- [10] M. Karplus, J. Am. Chem. Soc. 1963, 85, 2870-2871.
- [11] The carbon chemical shifts around the cyclopentane ring in 1 coincided with those of known 9,11-secosteroids. For details, see the Supporting Information.
- [12] Selected reviews for natural secosteroids: a) D. Sica, D. Musumeci, Steroids 2004, 69, 743-756; b) K. Penov Gasi, M. Sakac, S. Jovanovic-Santa, E. Djurendic, Curr. Org. Chem. 2014, 18, 216-259.
- [13] Selected reviews for ECD spectrum calculations and their applications for natural products: a) A. E. Nugroho, H. Morita, J. Nat. Med. 2014, 68, 1–10; b) G. Pescitelli, T. Kurtán, U. Flörke, K. Krohn, Chirality 2009, 21, E181–E201.
- [14] M. Cossi, N. Rega, G. Scalmani, V. Barone, J. Comput. Chem. 2003, 24, 669-681.
- [15] I. Ohtani, T. Kusumi, Y. Kashman, H. Kakisawa, J. Am. Chem. Soc. 1991, 113, 4092–4096.
- [16] Y. Yamaguchi, Y. Nakanishi, T. Shimokawa, S. Hashiguchi, A. Hayashi, Chem. Lett. 1992, 1713–1714.
- [17] L. A. Paquette, J. E. Hofferberth, Org. React. 2003, 62, 477 567.

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